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Phenolic compounds from the aqueous extract of Acacia catechu

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Two new phenolic compounds, 5-hydroxy-2-[2-(4-hydroxyphenyl) acetyl]-3-methoxylbenzoic acid (1) and (2S,3S)-3,7,8,3',4'-pentahydroxyflavane (2), were obtained from the aqueous extract of *Acacia catechu*, along with four known compounds identified as rhamnetin (3), 4-hydroxyphenyl ethanol (4), 3,3',5,5',7-pentahydroxyflavane (5), and fisetinidol (6). Their structures were determined on the basis of spectroscopic analysis. Free radical-scavenging activities of the new compounds were evaluated.

Keywords: Leguminosae; *Acacia catechu*; phenolic compounds; free radicalscavenging activity

1. Introduction

Acacia is the second largest genus in the Leguminosae family, comprising more than 1200 species worldwide, which are found in almost all habitats. The extract of Acacia catechu has been used in traditional Chinese medicine as antipyretic, anthelmintic, antipruritic, digestive, and antivenom agent. Preliminary pharmacological studies demonstrated antianemic, antiinflammatory, antimicrobial, anticancer, antioxidant, and antidiabetic activities [1,2]. Previous studies on this plant revealed the presence of flavanes [3,4], flavonoids [4], fatty acids [5], and gums [5]. In the course of our investigation on the bioactive constituents of A. catechu, two new phenolic compounds were obtained from A. catechu (Figure 1). In this paper, we present their isolation, structure elucidation, and biological evaluation of free radical-scavenging activities.

2. Results and discussion

Compound 1, obtained as green oily material, was assigned the molecular formula of C₁₆H₁₄O₆ by the pseudomolecular ion in HR-ESI-MS at m/z $301.0697 [M - H]^{-}$, with 10 degrees of unsaturation. The UV spectrum of 1 exhibited absorption maxima at 291 (3.99) and 202 (4.28) nm. Its IR spectrum showed the absorption bands for hydroxyl (3201 cm^{-1}) , conjugated carbonyl (1685 cm^{-1}), and aromatic ring (1600and 1515 cm^{-1}). In the ¹H NMR spectrum (Table 1), the AA'BB' system at δ 7.02 (2H, d, J = 8.4 Hz, H-2', 6') and 6.62 (2H, d)d, J = 8.4 Hz, H-3', 5') indicated the presence of a 1,4-disubstituted benzene ring. Two *meta*-coupled protons at δ 5.93 (1H, d, J = 1.2 Hz, H-6) and 5.92 (1H, d, J)J = 1.2 Hz, H-4) implicated the existence of a 1,2,3,5-tetrasubstituted benzene ring. In addition, one methoxyl group

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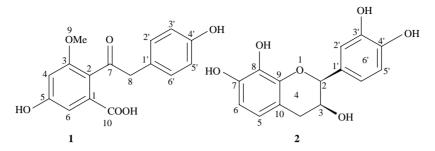


Figure 1. Structures of compounds 1 and 2.

Compound no.	1		2	
	$\delta_{ m H}$	δ_{C}	$\delta_{ m H}$	$\delta_{\rm C}$
1		103.3		
2		106.3	4.93 (m)	79.9
3		160.6	4.15 (m)	67.2
4	5.92 (d, <i>J</i> = 1.2)	93.3	3.11 (dd, J = 16.5, 4.5) 2.72 (dd, $J = 16.5, 3.3$)	34.0
5		169.0	6.41 (d, $J = 8.4$)	115.4
6	5.93 (d, $J = 1.2$)	91.7	6.38 (d, $J = 8.4$)	108.9
7		193.1		143.8
8	3.06 (s)	41.6		133.5
9	3.75 (s)	56.0		144.7
10		173.7		112.3
1'		125.7		132.0
2'	7.02 (d, $J = 8.4$)	132.4	7.04 (d, $J = 1.5$)	120.3
3'	6.62 (d, $J = 8.4$)	115.4		145.0
4'		156.9		145.4
5'	6.62 (d, $J = 8.4$)	115.4	6.77 (d, $J = 8.1$)	115.5
6'	7.02 (d, $J = 8.4$)	132.4	6.83 (d, $J = 8.1, 1.5$)	119.5

Table 1. ¹H and ¹³C NMR spectral data of **1** and **2** in CD_3COCD_3 (*J* in Hz).

resonating at δ 3.75 and one methylene at δ 3.06 (2H, H-8) were also displayed. In NOE difference spectrum, only the signal at δ 5.92 (H-4) was enhanced when irradiating the methoxyl, suggesting that the methoxyl group was located at C-3 (δ 160.6). The attachment of carboxyl group to C-1 was established by the HMBC correlations of H-4 (δ 5.92)/C-3, C-5 (δ 169.0) and C-6 (δ 91.7), H-6 (δ 5.93)/C-1 (δ 103.3), C-4 (δ 93.3), COOH (δ 173.7). The HMBC correlations of H-2' (δ 7.02)/C-8 (δ 41.6) and H-6' (δ 7.02)/C-8 indicated that B-ring was connected at C-8. The HMBC cross-peaks of H-8 (δ 3.06)/C-2 (δ 106.3) and H-8/C-7 (δ 193.1) indicated that 2-(4-hydroxyphenyl) acetyl was linked to C-2. Thus, compound **1** was elucidated as 5-hydroxy-2-[2-(4-hydroxy-phenyl)acetyl]-3-methoxybenzoic acid (Figure 2).

Compound **2** was isolated as colloid with $[\alpha]_D^{20} + 6.2$ (c = 0.5, MeOH). The molecular formula of $C_{15}H_{14}O_6$ was determined based on the quasi-molecular ion peak at m/z 289.0714 [M – H]⁻ by negative HR-ESI-MS, with nine degrees of unsaturation. The UV spectrum of **2** exhibited absorption maxima at 281 (3.5) and 205 (4.6) nm. The IR spectrum of **2** displayed absorption bands at 3202 (hydroxyl), 1609, and 1515 (aromatic

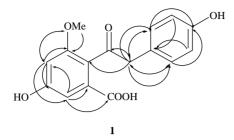


Figure 2. Key HMBC ($H \rightarrow C$) and NOESY (\leftrightarrow) correlations of **1**.

ring) cm⁻¹. The ¹H NMR spectrum of 2 (Table 1) displayed ABX-type aromatic protons at δ 7.04 (1H, d, J = 1.5 Hz, H-2'), 6.83 (1H, dd, J = 8.1, 1.5 Hz, H-6'), and 6.77 (1H, d, J = 8.1 Hz, H-5'), two orthocoupled aromatic protons at δ 6.41 (1H, d, $J = 8.4 \,\text{Hz}, \text{H-5}$) and 6.38 (1H, d, J = 8.4 Hz, H-6), two oxygenated methine protons at δ 4.93 (1H, m, H-2) and 4.15 (1H, m, H-3), one methylene at δ 3.11 (1H, dd, J = 16.5, 4.5 Hz, H-4a) and 2.72 (1H, dd, J = 16.5, 3.3 Hz, H-4b). The ¹³C NMR spectral data (Table 1) showed 15 carbons. The above NMR spectra of compound 2 were very similar to those of mesquitol [6,7], except that the two oxygenated methine protons were at δ 4.61 (1H, d, J = 7.2 Hz, H-2) and 3.99 (1H, m, H-3) and the chemical shifts of C-2 and C-3 were at δ 83.1 and 68.3, respectively, in mesquitol. The absolute configuration at C-3 was determined as S by comparing its CD spectral data with that in Ref. [8]. In the CD spectrum of 2, a positive Cotton effect at 350 nm was shown. Owing to the cis-configuration between H-2 and H-3, the absolute configuration of C-2 was also assigned to be S. Thus, the final structure of **2** was determined to be (2S,3S)-3,3',4',7,8-pentahydroxyflavane.

In addition, four known compounds were identified by comparing their spectroscopic data with those reported in the literature as rhamnetin (3) [9], 4-hydro-xyphenylethanol (4) [10], 3,3',5,5',7-pentahydroxyflavane (5) [11], and fisetinidol (6) [12].

New compounds 1 and 2 were evaluated *in vitro* for scavenging effects on ROS (HO, O₂, ROO, and H₂O₂), RNS (ONOO⁻), and 2,2-diphenyl-1-picrylhydrazyl radical (DPPH). There were significant differences between sample groups and model group. As far as the scavenging effect of stable DPPH was concerned, compound 2 exhibited the highest DPPH scavenging activity with absorbance values of 1.2966 \pm 0.1088 by comparing with the model group with absorbance values of 1.5769 \pm 0.054.

3. Experimental

3.1 General experimental procedures

The optical rotation values were determined with a JASCO P-2000 polarimeter. UV spectra were obtained with a JASCO V-650 spectrophotometer. IR spectra were obtained on an IMPACT 400 spectrometer. The ¹H NMR (300 MHz) and ¹³C NMR (100 MHz) spectra were recorded on a Mercury-300 and Mercury-400 spectrometers with TMS as an internal standard. NOE difference, HSQC, and HMBC spectra were run on Mercury-400 spectrometer. ESI-MS and HR-ESI-MS data were obtained from an Agilent 1100 LC/MSD Trap-SL mass spectrometer. CD spectra were measured on a JASCO J-810 spectropolarimeter. Silica gel GF₂₅₄ prepared for TLC and silica gel (160-200 mesh) for column chromatography (CC) were obtained from Qingdao Marine Chemical Factory (Qingdao, China). RP-C₁₈ silica gel was purposed from Merck Chemical Company Ltd. (Shanghai, China). HPLC separations were performed on a preparation YMC-Pack ODS-A column (10 μm, $250 \times 20 \,\mathrm{mm}$ ID) equipped with a Shimadzu SPD-6A UV spectrophotometric detector and a Thermo constametric pumping system. All the reagents were HPLC grade purchased from Beijing Chemical Company (Beijing, China).

3.2 Plant material

The aqueous extract of *A. catechu* was purchased from Beijing Pharmacy in July 2004 (Production Lot 20040309), and identified by Professor Lin Ma of the Institute of Materia Medica, Peking Union Medical College. A voucher specimen (ID-S-2285) has been deposited at the Herbarium of the Institute of Materia Medica, Chinese Academy of Medical Sciences & Peking Union Medical College, China.

3.3 Extraction and isolation

The medicine of A. catechu (1.0 kg) was extracted with water (5 L) using ultrasound for 45 min. The mixture was filtered and partitioned with EtOAc $(5L \times 8)$ and H₂O-saturated *n*-BuOH (5 L \times 8) successively. The EtOAc extract (200 g) was separated on a silica gel with CHCl₃-MeOH gradient solvent system (1:0 to 0:1) to obtain eight fractions (E1-E8), which were combined according to TLC analysis. Purification of E1 (180 mg) was performed by C-18 open CC eluting with a gradient system (MeOH-H₂O, 5:95 to 100:0) and led to the isolation of compound 4 (3 mg). Fraction E2 (5g) was subjected to silica gel CC eluting with petroleum etheracetone (9:1 to 1:1) to yield fractions E21–E28. The fraction of E22 (84 mg) was further purified by silica gel CC eluted with petroleum ether-acetone (9:1 to 1:1)to give compound 1 (4 mg). E26 (150 mg) was purified by C-18 CC eluted with MeOH $-H_2O$ (9:1 to 10:0), and was separated by RP-HPLC with MeOH- H_2O (1:1) to afford compound 3 (13 mg, $t_{\rm R}$ 18 min). Fraction E3 (50 g) was separated by C-18 open CC eluted with a gradient system (MeOH-H₂O, 5:95 to 100:0) to get fractions E31-E40. E32 (50 mg) was purified by reversed-phase HPLC eluting with MeOH-H₂O (30:70) to afford compounds 5 (3 mg, $t_{\rm R}$ 5 min) and **6** (6 mg, $t_{\rm R}$ 10 min). E36 (60 mg) was subjected to RP-HPLC eluted with MeOH-H₂O (30:70) to afford compound **2** (15 mg, t_R 10 min).

3.3.1 5-Hydroxy-2-[2-(4-hydroxyphenyl) acetyl]-3-methoxybenzoic acid (1)

Green oil, UV (MeOH) (nm) λ_{max} (log ε): 202 (4.28), 291 (3.99); IR (neat) ν_{max}^{KBr} (cm⁻¹): 3201, 1685, 1600, 1512; ¹H and ¹³C NMR spectral data, see Table 1; HR-ESI-MS *m*/*z*: 301.0696 [M - H]⁻ (calcd for C₁₆H₁₃O₆, 301.0712).

3.3.2 (2S,3S)-*3*,7,8,3',4'-Pentahydroxyflavane (2)

Colloid, $[\alpha]_D^{20}$ +6.2 (*c* 0.50, MeOH); UV (MeOH) (nm) λ_{max} (log ε): 205 (4.58), 281 (3.47); IR (neat) ν_{max}^{KBr} (cm⁻¹): 3202, 1609, 1600, 1515; CD (CH₂Cl₂:MeOH = 3:1): $\Delta \varepsilon_{350nm}$ +0.42; ¹H and ¹³C NMR spectral data, see Table 1; HR-ESI-MS *m/z* 289.0714 [M - H]⁻ (calcd for C₁₅H₁₃O₆, 289.0718).

3.4 Radical-scavenging activity assay

The antiradical activities of new compounds 1 and 2 were determined by the radical scavenging method described by Almeida et al. [13].

Acknowledgements

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